

# Development of Internet Technology TIPHAM (Tool for identity of Powdered Herbals through Analytical Microscopy) for Microscopic Identification of Crude Herbal Drugs

Dharya Singh, Vidhu Aeri, D. B. Ananthanarayana

Department of Pharmacognosy and Phytochemistry, School of Pharmaceutical Education and Research, New Delhi, India

Submitted: 30-11-2017

Revised: 10-01-2018

Published: ???

## ABSTRACT

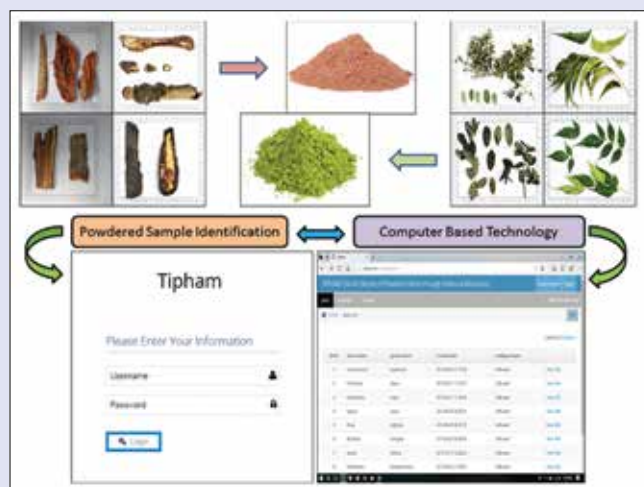
**Background:** Industrial usage of herbal plants has gone up, but techniques for verifying their botanical identity is still questionable. In the herbal industry, bulk consignments are received in powdered form as it is cumbersome to transport drugs in whole form. To ensure that the final product is safe and efficacious, the authenticity of the herbal plant should be established at the first stage. A proper methodology should be adopted in terms of computer technology to establish the correct botanical identity of the plant and to check the presence of substitutes and adulterants. **Objective:** To develop a software for identification of powdered samples of leaves and barks used in Ayurvedic Pharmacopoeia of India along with their substitutes and adulterants. **Materials and Methods:** Almost 100 plants have been selected from the Ayurvedic Pharmacopoeia of India comprising 54 barks and 46 leaves. Samples were self-collected and authenticated from the National Institute of Science Communication and Information Resources, Pusa, New Delhi. The selected crude herbal drugs were subjected to a detailed powdered microscopic identification and standard operating procedure for the preparation of slides was prepared. The features selected for identification of bark included 14 specific characters - stone cells, calcium oxalate crystals, starch grains, medullary rays, fibers, sclereids, cork, isolated oil cells, tubular lactiferous canals, phloem parenchyma, masses, rhytidoma, parenchyma, and secretory canals. These characters are further differentiated into 75 features and 151 subfeatures, whereas for leaves, 13 specific characters were included, namely, epidermis, stomata, trichomes, calcium oxalate crystals, fibers, cell contents, cystoliths, lamina, starch grains, tracheids, lactiferous canals, and xylem vessels which are differentiated into 139 features. The details of all the features have been uploaded in the software under the name tool for identity of powdered herbals through analytical microscopy (www.tiphm.com) with the database of 100 selected drugs. **Results:** A computer-based approach is developed which contains standard requirements for powdered plant parts, thus enabling identification of a bark or leaf powder in short time with minimum expertise. **Conclusion:** Computer-based technology would be a landmark in the field of pharmacognosy as proper identification of plant is the key to develop quality herbal products ensuring their safety and efficacy. **Key words:** Bark, computer based, pharmacognosy, tool for identity of powdered herbals through analytical microscopy leaf powder microscopy

## SUMMARY

Development of Internet Technology tool for identity of powdered herbals through analytical microscopy (TIPHAM) for microscopic identification of crude herbal drugs

- Samples of about 100 plants were self-collected from the National Institute of Science Communication and Information Resources. These samples were subjected to detailed powder microscopic evaluation with an aim to establish key diagnostic features to differentiate between powdered bark and leaf crude herbal drugs along with their substitutes and adulterants

- The features selected for identification of bark included 14 specific characters which are further differentiated into 75 features and 151 subfeatures, whereas for leaves, 13 specific characters which are classified into 139 features
- The details of all the features have been uploaded in the software under the name TIPHAM which contains database of 100 selected plants
- A computer-based approach is developed which will provide botanical authentication of powdered sample of bark or leaf in short time with minimum expertise.



**Abbreviations used:**  $\mu\text{m}$ : Micrometer; AHP: American Herbal Pharmacopoeia; DNA: Deoxyribonucleic acid; GMP: Good Manufacturing Practices; ICMR: Indian Council of Medical Research; Id: Identity Document; IT: Information Technology; MICROAID: Microaided Identification; MP: Megapixel; NA: Not Applicable; NISCAIR: National Institute of Science Communication and Information Resources; TIPHAM: Tool for Identity of Powdered Herbals through Analytical Microscopy; TLC: Thin-Layer Chromatography; UK: United Kingdom; WHO: World Health Organization.

## Correspondence:

Dr. Vidhu Aeri,  
Department of Pharmacognosy and  
Phytochemistry, School of Pharmaceutical  
Education and Research, Jamia Hamdard,  
New Delhi - 110 062, India.  
E-mail: vidhuaeri@yahoo.com  
DOI: 10.4103/pm.pm\_563\_17

## Access this article online

Website: [www.phcog.com](http://www.phcog.com)

## Quick Response Code:



This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: [reprints@medknow.com](mailto:reprints@medknow.com)

**Cite this article as:** Singh D, Aeri V, Ananthanarayana DB. Development of Internet Technology TIPHAM (Tool for identity of Powdered Herbals through Analytical Microscopy) for microscopic identification of crude herbal drugs. Phcog Mag 2018;XX:XX-XX.



Figure 1: Samples procured from industry. Industrial grounded powdered samples



Figure 2: Program microaided identification available in the United Kingdom. It consists of characters of all plant parts irrespective of leaf, bark, fruit, or seed. For example, trichomes and stomata are present in leaves; stone cells, vessels/tracheids, lignified parenchyma in barks; aleurone in seeds; pollen in fruits, whereas calcium oxalate crystals and starch grains are present in almost every plant part



Figure 3: Login page for tool for identity of powdered herbs through analytical microscopy program. Enter the user identity document and password and get access into the program



Figure 4: Homepage for tool for identity of powdered herbs through analytical microscopy program. Indicates logout option



Figure 5: Template for identification of a leaf powder Go to the option "Evaluate." Select leaf template. Features for identification of a leaf powder will appear. Enter the desired features and click "Search"

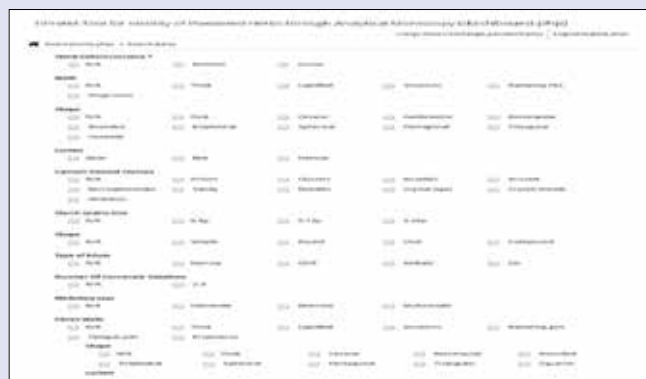
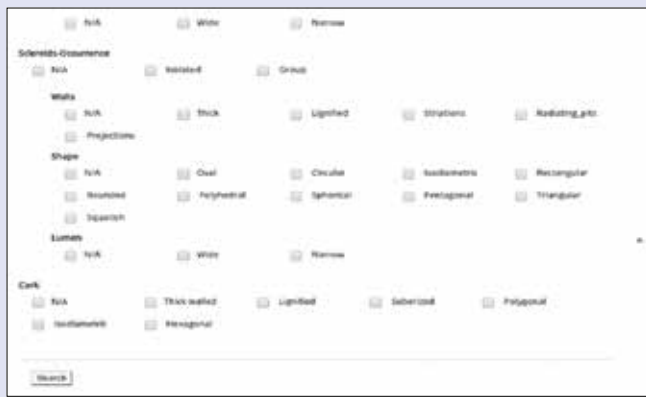


Figure 6: Template for identification of a bark powder. Go to the option "Evaluate." Select bark template. Features for identification of a bark powder will appear. Enter the desired features and click "Search"

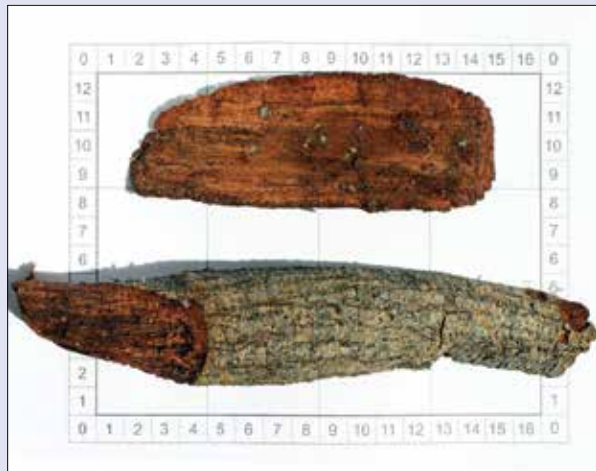
## INTRODUCTION

Herbal drugs have been used as dietary supplements, nutraceuticals, food additives, as ingredients in cosmetics, flavoring agents, and as fragrance. The industrial usage of plants and their parts, either as such

or after processing has gone up, the technology for confirming their botanical identity is still questionable. The knowledge of microscopical features is not known in industry. Therefore, the use of microscopic technique is coming down. It is still a powerful tool to identify powdered crude drugs. The herbal industry is generally procuring crude drugs in the powdered form as a convenient form to whole drugs. The fear of substitution and adulteration of raw herbs is also on rise as most of

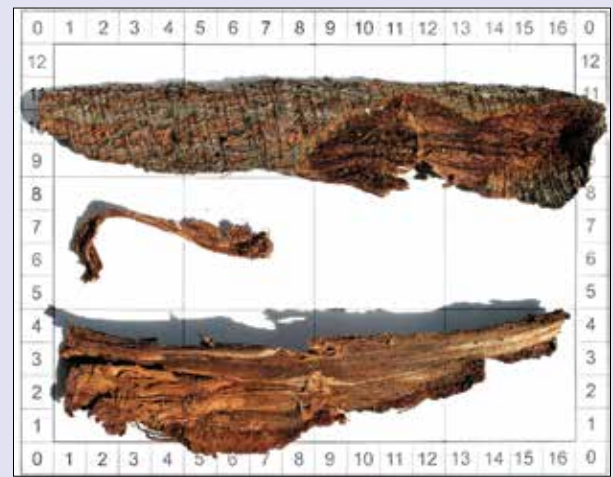


**Figure 7:** Template for identification of a bark powder. Go to the option "Evaluate." Select bark template. Features for identification of a bark powder will appear. Enter the desired features and click "Search"



**Figure 9:** Macroscopic characters of *Ficus racemosa* bark. Shape – Flat, curved, or channeled, Outer bark – Rough, whitish papery flakes coming out of outer surface, Inner bark – Pale brown, uneven, longitudinally striated, Fracture – Fibrous, Odor – Nil

the times only plant parts are available in the industry: 50% roots, 15% seeds, 12% wood waste, 9% whole plants, 7% bark/stem, 4% leaves, and 3% flowers are used as raw material. Performing macroscopy and microscopy of each plant part in every consignment in industry is a tedious, time-consuming job and one does not see good documentation leaving doubts about the quality control of the tests being done. Nonpharmacognosists are being trained to identify drugs, but they are unable to identify herbal plants due to nonavailability of data on microscopical features of crude powdered drugs. The pharmacopeias have introduced mandatory thin-layer chromatography testing of the plant material under examination. This has improved the identity testing subject to availability of specific marker compounds but cannot replace the botanical identity testing. Therefore, a computer-based technology needs to be developed to establish the correct botanical identity of the plant and check the presence of adulterants and allied drugs. A computer-based approach within seconds will help to provide the fingerprint of any part of the plant: leaves, seeds, fruits, bark, root, and stem. Such a technology would be a landmark in the field of pharmacognosy as proper identification of plant part is the key to develop a formulation of utmost quality and safety.<sup>[1]</sup>



**Figure 8:** Macroscopic characters of *Ficus lacor* bark. Shape – Flat-to-curved pieces, Outer bark – Ash to whitish gray, numerous transversely arranged lenticels, Inner bark – Reddish brown, rough, fibrous, and longitudinally striated, Fracture – Fibrous, Odor – Characteristic



**Figure 10:** Macroscopic characters of *Ficus religiosa* bark. Shape – Lat or slightly curved pieces, Outer bark – Uneven surface light brown to ash colored, Inner bark – Smooth and brownish, Fracture – Fibrous, Odor – Indistinct

The manufacture, sale, and distribution of herbal products are regulated in India under Drugs and Cosmetics Act 1940 and Rules 1945. The World Health Organization and other organizations emphasize the need for quality and standardization of plants used in manufacturing of traditional medicines including Ayurveda where the first basic requirement is establishing the correct botanical identity of plant drug attributing it to a specific genus and species.<sup>[2-5]</sup> In India, Ayurvedic Pharmacopoeia, Indian Pharmacopoeia, Indian Herbal Pharmacopoeia, and ICMR monographs have standards for checking authenticity of botanicals used in herbal products. However, serious efforts are not made to resolve the issue of controversial nomenclature.<sup>[6-8]</sup> The long history of safe usage of herbal medicines can be extrapolated only when the botanical identity of the plant going into those medicines is established and standardized.<sup>[9]</sup>

Microscopic identification is the most commonly used method for authentication of herbal drugs. Microscopic techniques examine structural and cellular features of herbs to determine their botanical origin.<sup>[10]</sup> This method is useful for identifying species with similar



**Table 1:** Microscopic characters bark powder

Stone cells				Sclereids			
Occurrence	Walls	Shape	Lumen	Occurrence	Walls	Shape	Lumen
N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
Isolated	Thick walled	Oval	Wide	Isolated	Thick walled	Oval	Wide
Group	Lignified	Circular	Narrow	Group	Lignified	Circular	Narrow
Abundant	Striations	Isodiametric	Pitted	Abundant	Striations	Isodiametric	Pitted
Few	Radiating pits	Rectangular	Slit like	Few	Radiating pits	Rectangular	
	Projections	Rounded			Projections	Rounded	
	Crenate margin	Polyhedral			Peg-like extension	Polyhedral	
	Beaded	Spherical			Beaded	Spherical	
	Pitted	Pentagonal			Branched	Pentagonal	
	Transversely pitted	Triangular			Three-sided thickening	Triangular	
	Branching pits	Squarish			Distinct pits	Cylindrical	
	Distinctly pitted	Irregular			Simple pits	Elongated	
	Devoid of pits	Oblong			Pitted walls	Fiber like	
	Thin walled	Angular			Thin walled	Very long	
						Blunt end	
						Pointed ends	
						Oblong	
						Cone shaped	
Fibers				Starch grains			
Occurrence	Walls	Shape	Lumen	Occurrence	Shape	Type of hilum	Number of concentric striations
N/A	N/A	N/A	N/A	Abundant	N/A	N/A	N/A
Isolated	Thick-walled	Oval	Wide	Few	Simple	Narrow	2-3
Numerous	Lignified	Circular	Narrow		Round	Cleft	Distinct striations
Group	Striations	Rectangular	Uneven		Oval	Radiate	
Few	Radiating pits	Rounded	Highly pitted wide lumen		Compound	Slit	
	Oblique pits	Polyhedral	Slit like		Spherical	Eccentric	
	Projections	Spherical			Triangular	Centrally located hilum	
	Frequently broken	Pentagonal			Small		
	Occasionally twisted	Triangular					
	Beaded	Squarish					
	Nonlignified	Irregular margin					
	Septate	Very long					
	Crenate margin	Pointed apex					
	Smoothly/finely striated	Bent					
	Dentate margin						
	Faint transverse septa						
	Thin walled						
	Irregular swellings						
	Crystal fibers						
	Transverse pits						
	Bifurcating ends						
	Distinct pits						
	Slit like						
Calcium oxalate crystals	Medullary rays	Cork	Isolated oil cells	Tubular lactiferous canals	Phloem parenchyma	Masses	Rhytidoma
N/A	N/A	N/A	Ovoid	N/A	N/A	N/A	N/A
Prismatic	Uniseriate	Thick walled	N/A	Groups	Spherical	Granular masses	Irregular
Clusters	Biseriate	Lignified		Oval	Oval	Dark brown cells containing tannins	Rectangular
Rosettes	Multiseriate	Suberized		Spherical	Pitted	Brown gummy	

Contd...

**Table 1:** Contd...

Calcium oxalate crystals	Medullary rays	Cork	Isolated oil cells	Tubular lactiferous canals	Phloem parenchyma	Masses	Rhytidoma
Acicular	Tangentially cut	Polygonal		Isolated	Lignified	Circular	
Microsphenoidal	Radially-cut	Isodiametric				Large oval	
Sandy	Longitudinally-cut	Hexagonal					
Needles		Rectangular					
Crystal layer		Pentagonal					
Crystal sheath		Beaded walls					
Idioblasts		Pitted lumen					
Rod shaped		Oval					
		Elongated					
		Thin walled					
		Multilayered					
		Radially arranged					
		Irregular					

NA: Not available

**Table 2:** Microscopic characters leaf powder

Epidermis	Stomata	Trichomes	Calcium oxalate crystals	Fibers	Stone cells and sclereids	Cell contents
N/A	N/A	Abundant	N/A	Isolated	Lignified	Volatile oil
Thick	Anomocytic	Few	Prismatic	Group	Non lignified	Mucilage
Thin	Anisocytic	N/A	Clusters	N/A	Wide lumen	Tannin cell
Beaded	Paracytic	Simple	Rosettes	Thick walled	Narrow lumen	Yellowish pigment
Straight	Diacytic	Branched	Acicular	Thin walled	Rectangular	Fatty oil globule
Wavy	Tetracytic	Covering	Microsphenoidal	Longitudinally cut	Oval	Resin ducts
Polygonal	Cyclocytic	Warty	Sandy	Narrow lumen	Triangular	Brownish matter
Anticlinical walls	Hexacytic	Glandular	Needles	Wide lumen	Fibrous	
Striated cuticle	Actinocytic	Unicellular	Crystal layer	Slit-like pits		
Devoid of stomata	Sunken	Thick walled	Crystal sheath			
Slightly sinous walled		Sessile	Idioblasts			
Hexagonal		Multicellular	Rod shaped			
Lignified wall		Uniseriate	Diamond-shaped idioblasts			
Dome-shaped papillae		Biseriate				
Papillose cells		Short stalk unicellular				
Lignified pith cells		Stalk multicellular				
Rectangular		Head unicellular				
		Oval head multicellular				
		Shaggy wooly trichomes				
		Straight				
		Irregularly bent				
		Striated walls				
		Pedestal base				
		Stellate				
		Spear shaped				
		Twisted				
		Narrow neck				
		Oval inflated apical cell				
		Thread-like terminals				
		Oval bicellular head				
		Conical				
		Cystolithic				
		Bicellular cylindrical stalk				
		8-12 celled head				
		Cup-shaped head				
		Globular head				
		Pointed apex				
		Short				
		Curved				
		Cicatrix				
		Bent				
		Long				

Contd...

Table 2: Contd...

Epidermis	Stomata	Trichomes	Calcium oxalate crystals	Fibers	Stone cells and sclereids	Cell contents
		Hook shaped Swollen bases Sickle shaped				
Cystoliths	Lamina	Starch grains	Tracheids	Lactiferous canals	Xylem vessels	
Cigar shaped	Thick-walled cells	Few	Spiral	Elongated	Annular	
Longitudinally cut	Multilayered epidermis	Abundant	Annular pitted	Tubular	Spiral	
Warty	Palisade cells	Simple	Bordered pitted	Yellowish granular latex	Pitted	
Cylindrical	Spongy parenchyma	Spherical		Long septate fragments	Reticulate	
		Compound Oval Pear shaped			Longitudinally cut Crystal fibers	

NA: Not available

Table 3: Represents specific distinguishing characters of barks of four species of *Ficus*

Features	<i>Ficus lacor</i>	<i>Ficus racemosa</i>	<i>Ficus religiosa</i>	<i>Ficus benghalensis</i>
Common name	Pakar <sup>[25]</sup>	Gular <sup>[26]</sup>	Pipal <sup>[27]</sup>	Bargad <sup>[28]</sup>
Stone cells				
Occurrence	Abundant	Isolated	Few	Isolated
Walls	Striated Distinctly pitted	Groups Beaded	Thick walled Pitted Striations	Thick walled Radiating pits
Shape	Isodiametric Squarish Oblong Oval	Polygonal Spherical Squarish	Triangular Oval	Circular Oval
Lumen	Narrow Wide Pitted	Wide Pitted	Narrow	Narrow Radiating pits
Size	15-45 µm [Figure 18 and 19]	>60 µm [Figure 20 and 21]	18-73 µm [Figure 22]	<50 µm [Figure 23]
Sclereids				
Occurrence	NA	NA	Abundant Isolated Group	Few
Walls	NA	NA	Striated Peg-like extensions	Beaded
Shape	NA	NA	Rectangular Irregular	Rectangular Oval
Lumen	NA	NA	Narrow Wide	Very long Wide
Size	NA	NA	<100 µm [Figure 24]	Pitted >100 µm [Figure 25]

NA: Not available

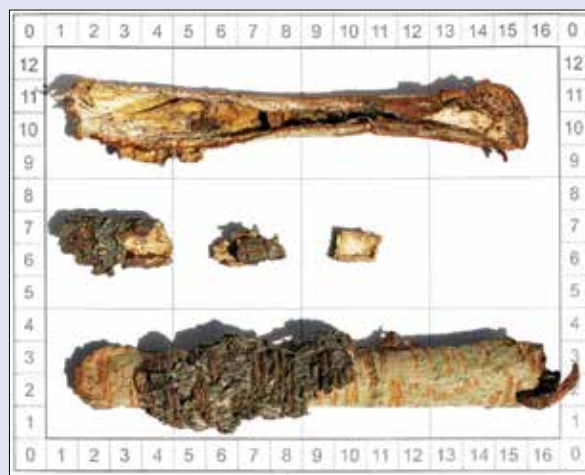
morphological characters. Today, there are a variety of methods available to authenticate herbal drugs, ranging from simple morphological examination to physical and chemical analysis and DNA molecular biology. Owing to cost, powder microscopy is still the most practical method for primary authentication.<sup>[11,12]</sup> The National and International Pharmacopoeia, namely, Chinese Materia Medica,<sup>[13]</sup> European Pharmacopoeia,<sup>[14]</sup> British Pharmacopoeia,<sup>[15]</sup> United States Pharmacopoeia,<sup>[16]</sup> Japanese Pharmacopoeia,<sup>[17]</sup> Ayurvedic Pharmacopoeia of India,<sup>[18]</sup> and Vietnamese Pharmacopoeia<sup>[19]</sup> emphatically provide the powder microscopic characters as one of the most identifying features of the crude herbal drug. Botanical microscopy is a unique, valuable, rapid, and cost-effective assessment tool. It continues to play an important role in the authentication and assessment of medicinal plants.

American Herbal Pharmacopoeia gives special emphasis on the powder microscopy of the herbal plants.<sup>[20]</sup>

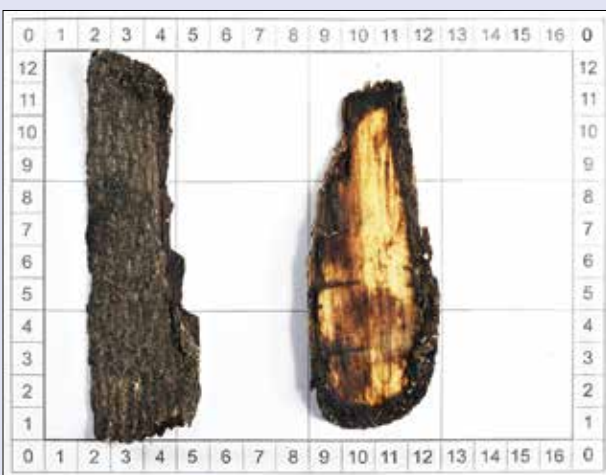
The aspect of identification and study of pharmacognostic properties of raw herbs appears to have not got the attention it deserves. Powdered drugs are used in herbal industry in formulations. To ensure that final product is safe and efficacious, its quality should be checked at first stage. Therefore, a databank should be developed which contain standard features of powdered plant parts. Individuals with minimal botanical and microscopical training can successfully identify powdered materials using this aid. This databank may also be modified to meet individual and industrial needs. The computer-based technology may reduce time and labor required to identify individual samples. It is known that computers are free from personal bias and will act according to



**Figure 11:** Macroscopic characters of *Ficus benghalensis* bark. Shape – Flat or somewhat curved, Outer bark – Ashy white color, transversely and longitudinally furrowed and cracked, Inner bark – Light brown in color, Fracture – Outer granular, inner hard, fibrous, and pinkish in color, Odor – Not characteristic



**Figure 12:** Macroscopic characters of *Albizia lebbeck* bark. Shape – Thick and flat pieces, Outer bark – Dark grayish brown, rough due to longitudinal and transverse cracks, Inner bark – Light yellow to gray and fibrous. Fracture – Laminated in outer region and fibrous in inner region, Odor – Characteristic



**Figure 13:** Macroscopic characters of *Albizia odoratissima* bark. Shape – Flat pieces, Outer bark – Dark blackish gray, Inner bark – Light yellowish brown and longitudinally striated, Fracture – Short and Fibrous, Odor – Characteristic



**Figure 14:** Macroscopic characters of *Azadirachta indica* leaf. Type of leaf – Compound, Shape – Lanceolate, Arrangement – Subopposite or alternate, Apex – Acute, Margin – Serrate, Venation – Reticulate, Base – Oblique, Texture – Glabrous, Color – Slightly yellow green, Odor – Indistinct

information fed into them. Hence, there are zero chances of error using this technology.<sup>[1]</sup>

## MATERIALS AND METHODS

### Selection of barks and leaves

About 53 barks and 46 leaves were selected from seven volumes of Ayurvedic Pharmacopoeia of India to study powder microscopy. Fresh bark and leaf samples were self-collected from the National Institute of Science Communication and Information Resources Pusa campus and Botanical Garden, Noida. These samples were identified by Dr. Sunita Garg, Scientist G.

### Preparation of database

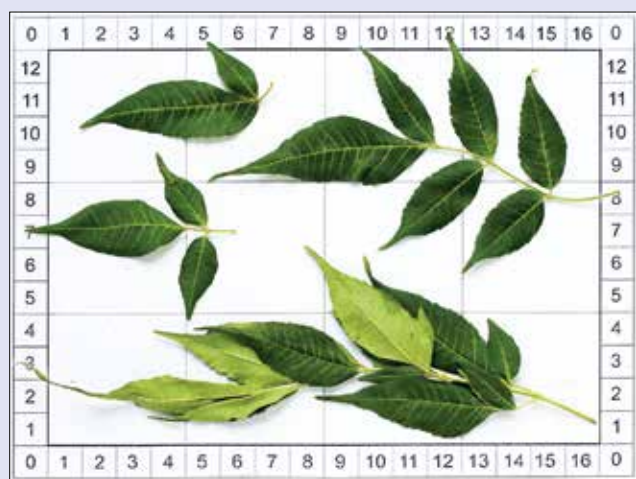
The selected barks and leaves were subjected to detailed powdered microscopic identification. The specific identifying features of each bark

**Table 4:** Represents specific distinguishing characters of barks of *Albizia lebbeck* and *Albizia odoratissima*

Features	<i>Albizia lebbeck</i>	<i>Albizia odoratissima</i>
Common name	Shiris <sup>[29-31]</sup>	Kala Shiris <sup>[32]</sup>
Calcium oxalate crystals	Prismatic [Figure 26]	Rosette [Figure 27]
Stone cells		
Occurrence	NA	Isolated Group
Walls	NA	Thin walled
Lumen	NA	Wide
Shape	NA	Oval [Figure 28]
Cork	Oval Rounded [Figure 29]	Thick walled Polygonal Pentagonal [Figure 30]

NA: Not available

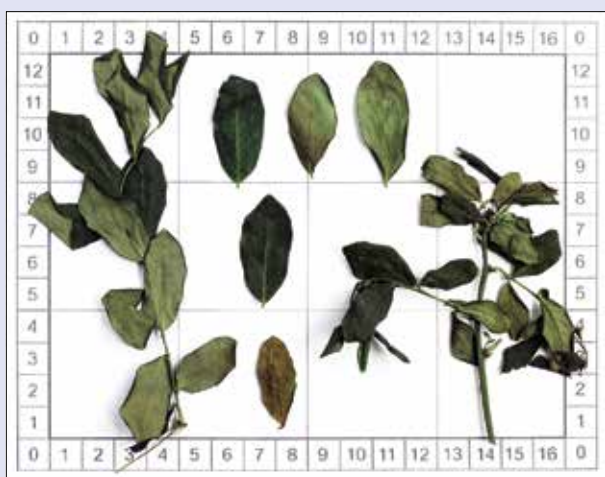




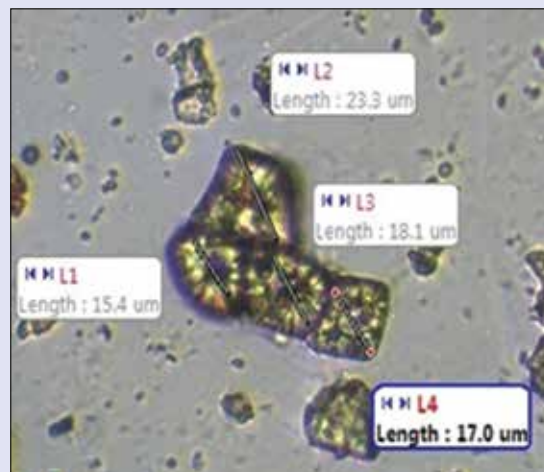
**Figure 15:** Macroscopic characters of *Melia azedarach* leaf. Type of leaf – Bipinnate, Shape – ovate to oblong lanceolate, Arrangement – More or less opposite, Apex – Acuminate, Margin – Entire to variously serrate, Venation – Reticulate, Base – Slightly unequalateral, acute, or rounded, Texture – Glabrescent, Color – Dark green, Odor – Pungent odor when crushed



**Figure 16:** Macroscopic characters of *Indigofera tinctoria* leaf. Type of leaf – Compound, Shape – Oblong to oblanceolate, Arrangement – Alternate, Apex – Rounded tip, Margin – Entire, Base – Rounded, Texture – Velvety, Color – Pale green to greenish black, Odor – No characteristic odor



**Figure 17:** Macroscopic characters of *Indigofera arrecta* leaf. Type of leaf – Bipinnate or tripinnate, Shape – Oblong, Arrangement – Opposite, Apex and base – Acute, Margin – Entire, Color – Dark greenish black



**Figure 18:** Stone cells present in *Ficus lacor* bark powder. Abundant stone cells with size ranging from 15-25  $\mu\text{m}$  in length

**Table 5:** Represents specific distinguishing characters of leaves of *Azadirachta indica* and *Melia azedarach*

Features	<i>Azadirachta indica</i>	<i>Melia azedarach</i>
Common name	Neem <sup>[33-36]</sup>	Bakayana <sup>[37]</sup>
Epidermis	Straight Anticlinical	Wavy
Trichomes	Simple Covering Warty [Figure 31]	Glandular [Figure 32] Stellate [Figure 33]
Stomata	Anomocytic Actinocytic [Figure 34]	Anomocytic [Figure 35]
Calcium oxalate crystals	Prismatic [Figure 36]	Needles [Figure 37]

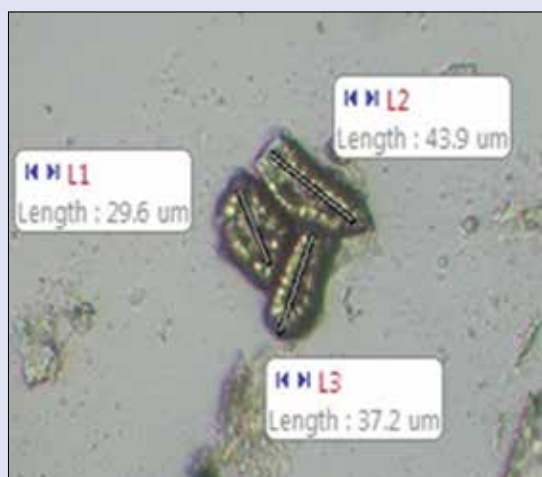
and leaf were studied along with few selected adulterants and substitutes. These features of 100 plants were uploaded in the software tool for

identity of powdered herbs through analytical microscopy (TIPHAM). The powder microscopy of bark samples of four *Ficus* species, that is, *Ficus lacor*, *Ficus racemosa*, *Ficus religiosa*, and *Ficus benghalensis* along with *Albizia lebbek* and *Albizia odoratissima* and leaves of *Azadirachta indica*, *Melia azedarach*, *Indigofera tinctoria*, and *Indigofera arrecta* is discussed in the result section as an example.

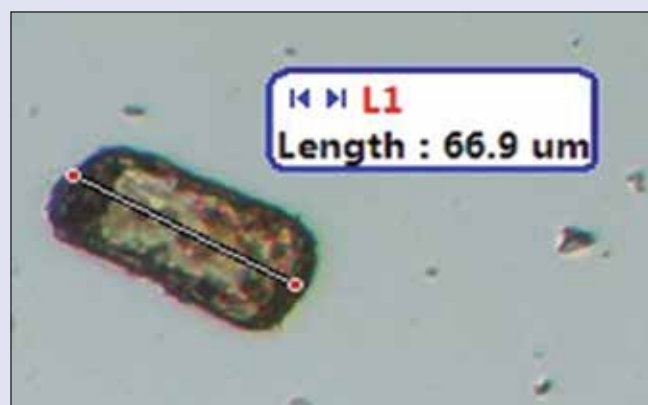
### Processing of crude drug

Standard operating procedure for preparation of slides of powdered bark and leaf samples was prepared with varying grinding techniques – industrial and laboratory blade grinding. Five samples were procured from industry [Figure 1] and microscopical features were compared using Motic microscope among various samples of these commonly used herbal drugs. It was concluded that grinding technique does not affect significantly probability of various microscopical features. The microscopical features were found to be stable and specific, which can be used to determine the botanical identity of the drug.<sup>[21]</sup>

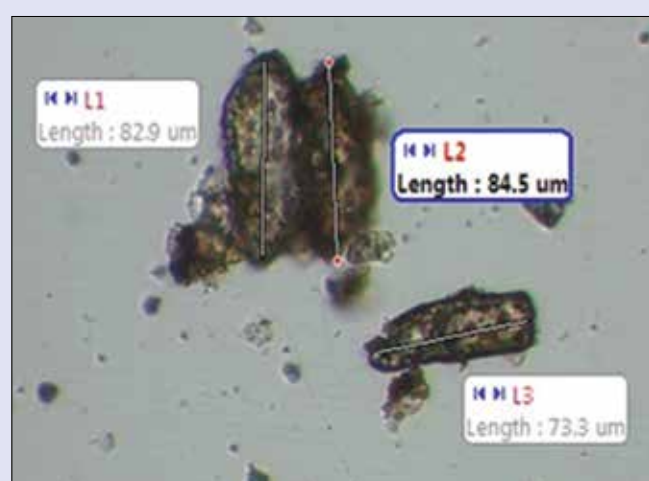




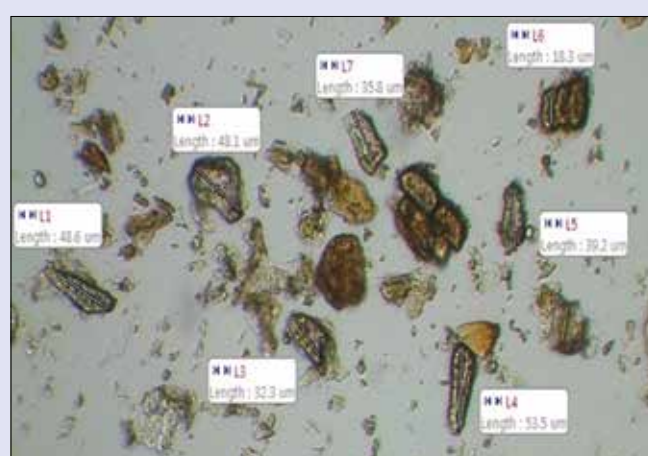
**Figure 19:** Stone cells present in *Ficus lacor* bark powder. Abundant stone cells with size ranging from 25-45  $\mu\text{m}$  in length



**Figure 20:** Stone cells present in *Ficus racemosa* bark powder. Stone cells larger in size than *Ficus lacor*, that is, greater 60  $\mu\text{m}$  in length



**Figure 21:** Stone cells present in *Ficus racemosa* bark powder. Stone cells larger in size than *Ficus lacor*, that is, greater 60  $\mu\text{m}$  in length



**Figure 22:** Stone cells present in *Ficus religiosa* bark powder. Few stone cells ranging 18-73  $\mu\text{m}$  in length

**Table 6:** Specific distinguishing characters of leaves of *Azadirachta indica* and *Melia azedarach*

Features	<i>Indigofera tinctoria</i>	<i>Indigofera arrecta</i>
Common name	Nili <sup>[38]</sup>	Bengal indigo <sup>[39]</sup>
Trichomes	Oval head Multicellular [Figure 38]	Simple Unicellular [Figure 39]
Calcium oxalate crystals	NA	Prismatic [Figure 40]

NA: Not available

### Standard operating procedure for preparation of slides

Two types of slides were prepared for the visualization of microscopical features present in a bark. The slide preparation method was optimized to determine the dilution of powdered bark and leaf in water to visualize common as well as distinguishing characters.

Slide I: A 500 mg of moderately fine (44/85) and fine (85) powdered material was soaked overnight in 10 ml of water (1:20) for 24 h. Subsequently, the contents were poured in a Petri plate and slide was

prepared by mounting the contents on a clean and dried slide with a brush and observed under Motic microscope moticam 3.0 MP, AE 2000. Most of the features were visible except stone cells and sclereids, which require treatment by oxidizing agent.

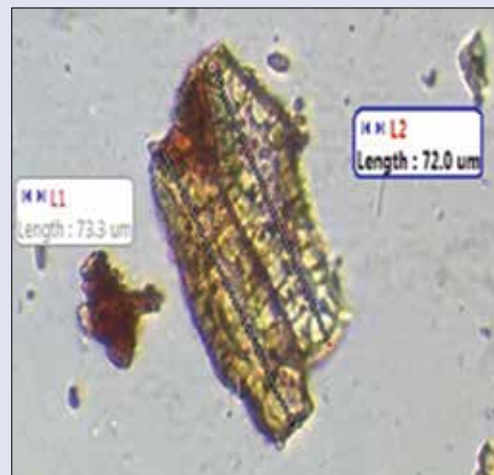
Slide II: The slide was prepared by potassium chlorate treatment which is used as an oxidizing agent used for disruption of stone cells and sclereids. However, calcium oxalate crystals and starch grains are destroyed using this method. A 200 mg of powdered material was boiled with 5 ml of 50% nitric acid. To this, added a pinch ~100-150 mg of potassium chlorate. The contents in a Petri plate are poured after the effervescence ceases. The contents were mounted on a clean and dried glass slide with the help of a brush; observed under Motic microscope.

### Development of parameters to study powder microscopy of crude drugs

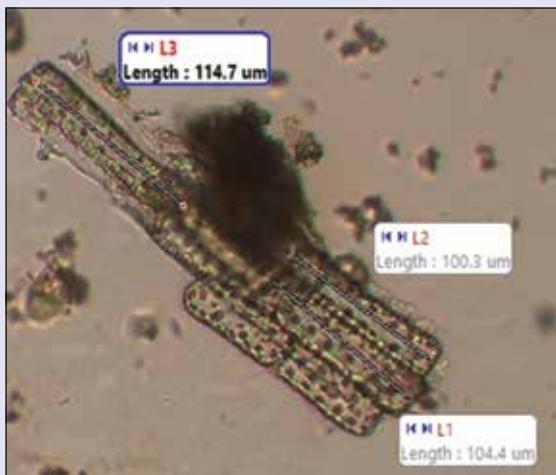
A bark powder is identified microscopically by various features such as stone cells, sclereids, calcium oxalate crystals, medullary rays, fibers, starch grains, and cork. Bark powder cannot be merely authenticated by observing the presence or absence of these features as these are common in all barks. The variation lies in the size and shape of the stone cells and sclereids, their occurrence, and type of wall and lumen. The program TIPHAM is developed keeping in mind the specificity of these parameters for a particular bark. Similar case stands for authentication of the leaf



**Figure 23:** Stone cells present in *Ficus benghalensis* bark powder. Isolated or group of stone cells about < 50 µm in length



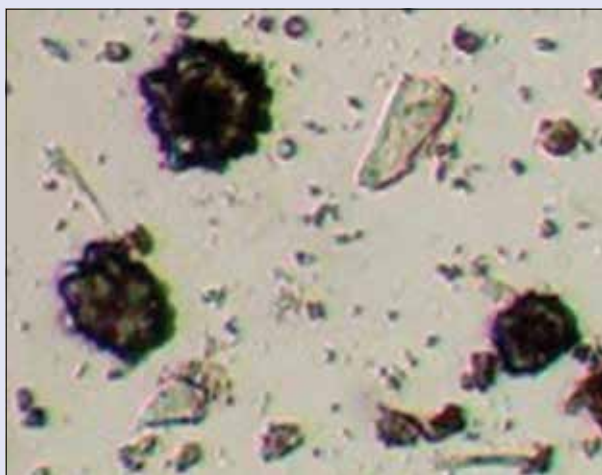
**Figure 24:** Sclereids present in *Ficus religiosa* bark powder. Abundantly present < 100 µm in length



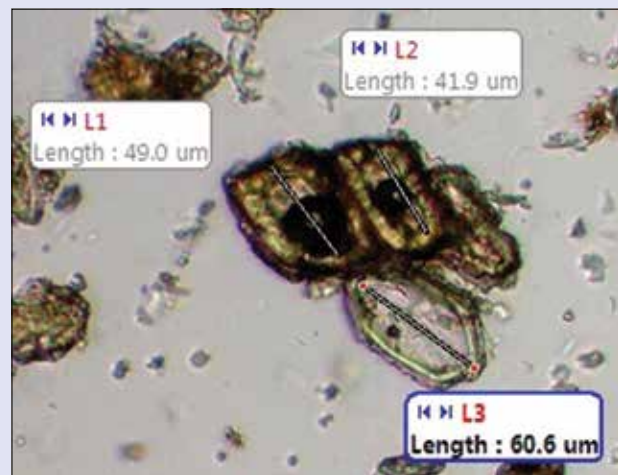
**Figure 25:** Sclereids present in *Ficus benghalensis* bark powder. Few in number > 100 µm in length



**Figure 26:** Prismatic crystals present in *Albizia lebbeck* bark powder. Prismatic crystals of calcium oxalate observed in *Albizia lebbeck* bark powder

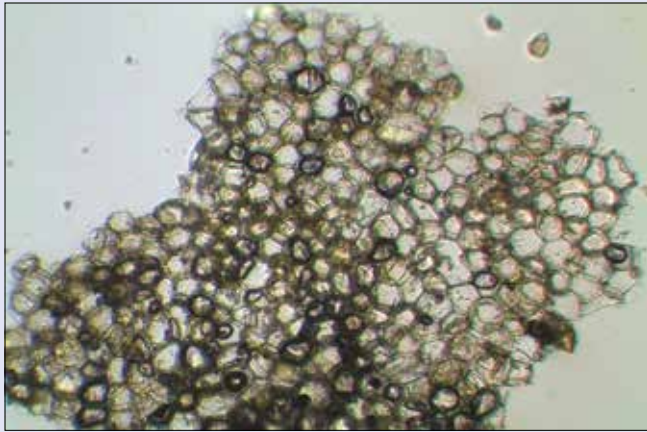


**Figure 27:** Rosette crystals present in *Albizia odoratissima* bark powder. Rosette crystals of calcium oxalate which is a distinguishing character from *Albizia lebbeck* bark powder



**Figure 28:** Stone cells present in *Albizia odoratissima* bark powder. Isolated or group of oval stone cells which are found to be absent in *Albizia lebbeck* bark powder





**Figure 29:** Cork cells present in *Albizia lebeck* bark powder. Oval-to-rounded cells due to disintegration of parenchymatous cells which is a characteristic feature of *Albizia lebeck* bark powder cavities in cork



**Figure 30:** Cork cells present in *Albizia odoratissima* bark powder. Polygonal-to-pentagonal cork cells



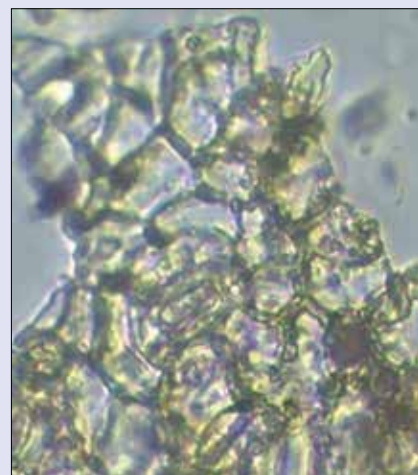
**Figure 31:** Trichomes present in *Azadirachta indica* leaf powder. Simple, covering, and warty trichomes



**Figure 32:** Trichomes present in *Melia azedarach* leaf powder. Glandular trichomes present which is a distinguishing character from *Azadirachta indica* leaf powder



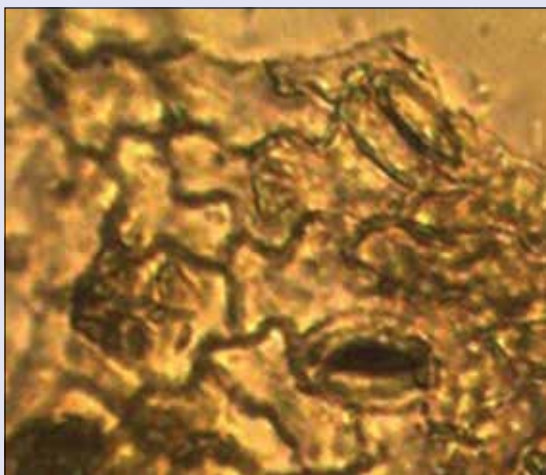
**Figure 33:** Trichomes present in *Melia azedarach* leaf powder. Stellate trichomes present which is a distinguishing character from *Azadirachta indica* leaf powder



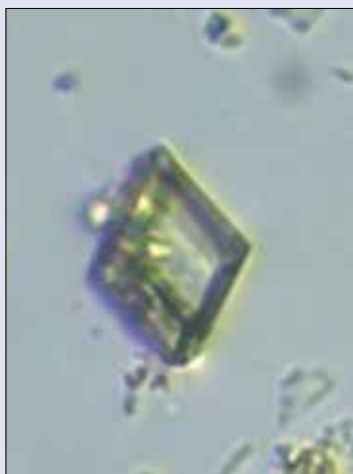
**Figure 34:** Stomata present in *Azadirachta indica* leaf powder. Anomocytic and actinocytic stomata present with straight and anticlinal epidermal walls

powder. The powder microscopy of bark was studied based on specific identifying features and their subfeatures as shown in Table 1. The

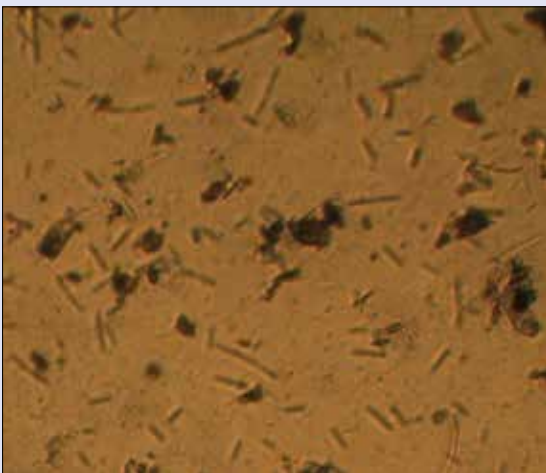




**Figure 35:** Stomata present in *Melia azedarach* leaf powder. Anomocytic stomata present with wavy epidermal walls



**Figure 36:** Calcium oxalate crystals present in *Azadirachta indica* leaf powder. Prismatic crystals of calcium oxalate



**Figure 37:** Calcium oxalate crystals present in *Melia azedarach* leaf powder. Needle-shaped crystals of calcium oxalate

powder microscopy of leaves was studied based on following parameters as shown in Table 2.

### Development of tool for identity of powdered herbs through analytical microscopy

TIPHAM was based on Microaided identification-1979 (MICROAID). The program MICROAID is available in the UK which contains databank of 174 powdered vegetable drugs.<sup>[22,23]</sup> Such software is not available in India. MICROAID consists of a common program which contains all the features of a medicinal plant irrespective of whether it is a bark, leaf, seed, or fruit [Figure 2], whereas in TIPHAM features have been segregated individually for the leaf and bark.

Our group has developed IT tool software on a pilot scale for identification of crude drugs. Database has been generated for about 100 crude drugs. Powdered samples were given to different volunteer students to check validity and application of software. Desirable results were obtained, but it needs to be further supported by feeding more data that can be achieved by working on other plant parts as well.

### Flow chart for tool for identity of powdered herbs through analytical microscopy

The web address ([www.tiphm.com](http://www.tiphm.com)) enables the user to visit the webpage. This webpage will require the user to fill the login id and password to use the software [Figure 3]. Subsequently, user will be taken to the homepage of the tool [Figure 4]. Once the user selects the "Home" option, templates for both bark and leaf will appear [Figures 5-7]. The specific identifying characters are entered in the bark or leaf template and click "Search." Software will search for that particular bark or leaf powder which contains similar features as entered from the database.

### Experimental procedure

All the chemicals used in the experiments were of analytical grade. Potassium chlorate was procured from Sigma-Aldrich. Potassium chlorate and 50% nitric acid form a liquid known as Schulze's maceration fluid and is used to disintegrate hard woody substances such as sclereids and stone cells. This fluid is a powerful oxidizing agent and it rapidly oxidizes and removes lignin from vegetable tissues.<sup>[24]</sup> Two types of slides were prepared – overnight soaking with water for 24 h and other by treatment with potassium chlorate.

### RESULTS

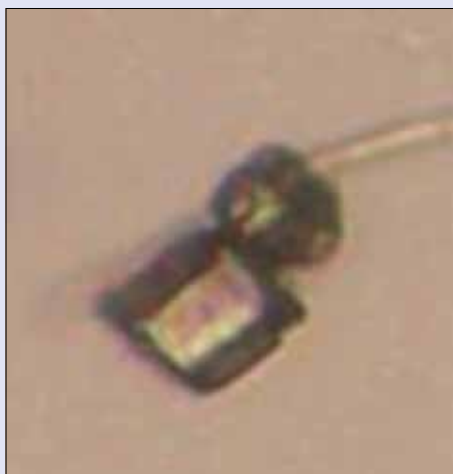
Microscopy on powdered samples of 54 barks and 46 leaves has been performed along with few selected adulterants and allied species. The given below is an example of powder microscopy of four bark species of *Ficus* and two species of *Albizia* along with leaves of *A. indica*, *M. azedarach*, *I. tinctoria*, and *I. arrecta*. These samples have been analyzed for specific distinguishing features. Figures 8-13 represent self-collected samples of barks of *F. lacor*, *F. racemosa*, *F. religiosa*, *F. benghalensis*, *A. lebeck*, and *A. odoratissima*, respectively, whereas Figures 14-17 represent self-collected samples of leaves of *A. indica*, *M. azedarach*, *I. tinctoria*, and *I. arrecta*, respectively. Tables 3 and 4 represents specific distinguishing characters of barks of four species of *Ficus* and *A. lebeck* and *A. odoratissima*, respectively, whereas Tables 5 and 6 indicate specific distinguishing characters of leaves of *A. indica*, *M. azedarach*, and *I. tinctoria*, *I. arrecta*, respectively. These specific identifying characters are entered in the bark template of TIPHAM and click "search." Software will search for that particular bark powder which possesses these features from the database. Hence, a bark or a leaf powder can be identified within seconds.



**Figure 38:** Trichomes present in *Indigofera tinctoria* leaf powder. Abundant multicellular trichomes with oval head



**Figure 39:** Trichomes present in *Indigofera arrecta* leaf powder. Simple and unicellular trichomes



**Figure 40:** Calcium oxalate crystals in *Indigofera arrecta* leaf powder. Prismatic crystals of calcium oxalate which were not observed in *Indigofera tinctoria* leaf powder

## DISCUSSION

The first step for quality control and authentication of any herbal drug is to study its morphology followed by its anatomy or microscopy. Powdering

of crude drugs in industries is a dust generating process. Ayurvedic herbal industries are also shifting toward good manufacturing practices. Therefore, industries are willing to purchase grounded herbs, which make it absolutely necessary to check its botanical identity at first stage. This software is beneficial from industrial standpoint because most of the herbal drugs supplied today are in powder form. Hence, there are more chances of adulteration as it is very easy to spoil a drug in powdered state. Industry emphasizes for powder microscopy as it is effortless. There is no need to cut sections and minimum equipment and expertise required. This software will facilitate to do the same. The botanical identity of any plant can be confirmed within seconds. Moreover, this software will be beneficial for pharmacopoeial laboratories.

Different bark powders are more or less similar in appearance. Based on the powder microscopic features, the bark powders of four *Ficus* species can be distinguished. *F. lacor* showed wide range of stone cells in terms of size and shape as compared to other three species of *Ficus*, whereas sclereids are found to be absent in *F. lacor* and *F. racemosa* but abundant in *F. religiosa* and few in *F. benghalensis*. *A. lebbek* bark powder showed prismatic crystals of calcium oxalate, devoid of stone cells, and contains cork cells possessing oval or rounded cavitations which is the most distinguishing character for this bark powder. On the other hand, *A. odoratissima* contains rosette calcium oxalate crystals, oval stone cells, and polygonal-to-pentagonal cork cells.

*A. indica* leaf powder showed straight epidermal walls embedded with anomocytic and actinocytic stomata; simple, covering, and warty trichomes along with prismatic crystals of calcium oxalate, whereas *M. azedarach* leaf powder showed wavy epidermal walls embedded with actinocytic stomata; glandular and stellate trichomes with needle-shaped crystals of calcium oxalate. *I. tinctoria* leaf powder revealed the presence of oval head multicellular trichomes, whereas *I. arrecta* possess simple unicellular trichomes.

These specific features when entered in the TIPHAM software, the tool will identify these distinguishing characters and reveal the botanical identity of that particular bark or leaf powder.

## CONCLUSION

Powder microscopy of bark and leaves has been performed and information is fed into TIPHAM program. The databank will also comprise of information about closely related species of a plant part and adulterants making the identification more effective. This IT tool software has been developed on a pilot scale for identification of about 100 crude drugs, but it needs to be further supplemented by incorporating more data on other plant parts as well.

## Acknowledgements

We thank Ayurvedye Trust, Bengaluru, which is a nonprofitable charitable trust for funding development of IT tool TIPHAM. We thank Dr. Sunita Garg, Scientist G, NISCAIR, Pusa, for her support in collection and identification of plants.

## Financial support and sponsorship

Nil.

## Conflicts of interest

There are no conflicts of interest.

## REFERENCES

1. Narayana DB. Pharmacognosy and technology. *Pharmacogn Mag* 2010;6:145-6.
2. World Health Organisation. *Quality Control Methods for Medicinal Plants*. Geneva: World Health Organisation; 1998.

3. World Health Organisation. Traditional Medicine Strategy. Geneva: World Health Organisation; 2002-2005.
4. European Medicine Evaluation Agency (EMA). Guidelines on Quality of Herbal Medicinal Products. EMA/CVMP/814/00. London: European Medicine Evaluation Agency; 2006.
5. World Health Organisation (WHO). Guidelines on Good Agricultural and Collection Practices (GACP) for Medicinal Plants. Geneva: World Health Organisation; 2003.
6. Warriar PK, Nambiar VP, Ramankutty C, editors. Indian Medicinal Plants. Vol. 1-5. India: Orient Black Swan/Universities Press; 1994.
7. Sivarajan VV, Balchandran I, editors. Ayurvedic Drugs and Their Plant Sources. New Delhi: Oxford and IBH Publishing Co. Pvt. Ltd.; 1994.
8. Dixit VK. Controversial ayurvedic herbs. *J Adv Pharm Technol Res* 2011;2:78-80.
9. Rajendra MD, Ananthanarayana DB. Controversial nomenclature of ayurvedic drugs: Challenges for scientists. *Phcog Rev* 2009;3:1-7.
10. Smillie TJ, Khan IA. A comprehensive approach to identifying and authenticating botanical products. *Clin Pharmacol Ther* 2010;87:175-86.
11. Torey A, Sasidharan S, Yeng C, Latha LY. Standardization of cassia spectabilis with respect to authenticity, assay and chemical constituent analysis. *Molecules* 2010;15:3411-20.
12. Zhao ZZ, Hu YN, Wong YW, Gigi Wong WC, Wu K, Jiang ZH, *et al.* Application of microscopy in authentication of Chinese patent medicine – Bo Ying compound. *Microsc Res Tech* 2005;67:305-11.
13. The Editorial Committee. *Materia Medica of China*. Shanghai: Shanghai Science and Technology Publishing House; 1999.
14. European Pharmacopoeia Commission. *European Pharmacopoeia*. 6<sup>th</sup> ed. Strasbourg: European Directorate for the Quality of Medicines and Healthcare of Council of Europe; 2007.
15. British Pharmacopoeia Commission. *British Pharmacopoeia* 2009. London: The Stationery Office on Behalf of the Medicines and Healthcare Products Regulatory Agency; 2008.
16. The United States Pharmacopoeial Convention. *The United States Pharmacopoeia*, 30<sup>th</sup> Revision/National Formulary. 25<sup>th</sup> ed. Rockville: The United States Pharmacopoeial Convention; 2005.
17. Japanese Pharmacopoeia Committee. *The Japanese Pharmacopoeia*. 15<sup>th</sup> ed. Tokyo: Society of Japanese Pharmacopoeia; 2006.
18. The Committee of the Ayurvedic Pharmacopoeia of India. *The Ayurvedic Pharmacopoeia of India*. New Delhi: The Controller of Publications Civil Lines; 2001.
19. Vietnamese Pharmacopoeia Commission, Ministry of Health. *Vietnamese Pharmacopoeia*. 3<sup>rd</sup>. Hanoi, Vietnam: Ministry of Health – S.R. Vietnam; 2005.
20. *American Herbal Pharmacopoeia*, Roy Upton Herbalist; April, 1999.
21. Singh D, Aeri V, Narayana DBA. Development of Standard Operating Protocol for Slide Preparation of Powdered Bark Samples with Varying Grinding Techniques. *Pharmacog J* 2018;10:265-71.
22. Jolliffe GH, Jolliffe GO. The microcomputer as an analytical aid in drug microscopy. In: Evans WC, editors. *Trease and Evans' Pharmacognosy*. 13<sup>th</sup> ed. London: Bailliere Tindall; 1989. p. 784-90.
23. Jolliffe GH, Jolliffe GO. Computer-aided identification of powdered vegetable drugs. *Analyst* 1976;101:622-33.
24. Chauhan MG. *Microscopic Profile of Powdered Drugs Used in Indian System of Medicine*, Volume 1, Bark Drugs. Jamnagar, Gujarat, India: Published by Institute of Ayurvedic Medicinal Plant Sciences; 2007. p. 124.
25. Pharmacopoeia Commission for Indian Medicine & Homoeopathy. *Ayurvedic Pharmacopoeia of India*. Part I. Vol. II. Ghaziabad, UP: Ministry of Health and welfare, Government of India. p. 144.
26. Gupta AK, Tandon N, Sharma M. *Quality Standards of Indian Medicinal Plants*. Vol. 9. New Delhi: Medicinal Plant Unit, ICMR; 2008. p. 167.
27. Koilpillai B, Sabesan GS, Rai S. Comparative pharmacognostic studies on the barks of four *Ficus* species. *Turk J Bot* 2010;34:215-24.
28. Gupta AK, Tandon N, Sharma M. *Quality Standards of Indian Medicinal Plants*. Vol. 7. New Delhi: Medicinal Plant Unit, ICMR; 2008. p. 117.
29. Pharmacopoeia Commission for Indian Medicine & Homoeopathy. *Ayurvedic Pharmacopoeia of India*, Part I. Vol. III. Ghaziabad: Pharmacopoeia Commission for Indian Medicine & Homoeopathy; 2016. p. 201.
30. Gupta AK, Tandon N, Sharma M. *Quality Standards of Indian Medicinal Plants*. Vol. 2. New Delhi: Medicinal Plant Unit, ICMR; 2008. p. 1.
31. Chahuhan M. *Microscopic Profile of Powdered Drugs Used in Indian System of Medicine*. Volume 1, Bark Drugs. Jamnagar, Gujarat, India: Published by Institute of Ayurvedic Medicinal Plant Sciences; 2007. p. 96.
32. Orwa C, Mutua A, Kindt R, Jamnadass R, Anthony S. *Agroforestry Database 4.0*; 2009. p. 1-5.
33. Faisal M, Sridhar B, Kumar Sunil KN, Sudhakar Ravi M. Macro microscopic fingerprints of panchanga of *Ishwari-Aristolochia indica* Linn. *J Phytopharmacol* 2015;4:61-7.
34. Government of India. *Ayurvedic Pharmacopoeia of India*. Part I. Vol. II. Ghaziabad: Ministry of Health and welfare, Government of India; 2001. p. 132.
35. Gupta AK, Tandon N, Sharma M. *Quality Standards of Indian Medicinal Plants*. Vol. 11. New Delhi: Medicinal Plant Unit, ICMR; 2008. p. 83.
36. Chahuhan M. *Microscopic Profile of Powdered Drugs Used in Indian System of Medicine*, Volume 2, Bark Drugs. Jamnagar, Gujarat, India: Published by Institute of Ayurvedic Medicinal Plant Sciences; 2007. p. 98.
37. Chahuhan M. *Microscopic Profile of Powdered Drugs Used in Indian System of Medicine*. Volume 2, Bark Drugs. Jamnagar, Gujarat, India: Published by Institute of Ayurvedic Medicinal Plant Sciences; 2007. p. 84.
38. Santhan P. Leaf structural characteristics of important medicinal plants. *Int J Res Ayurveda Pharm* 2010;5:673-9.
39. Suvarnalatha A, Yasodamma N, Alekhya C, Chaithra D. Pharmacognostic studies of *Indigofera hirsuta* L. *Int J Pharm Pharm Sci* 2014;6:111-7.